Effectiveness of Mouthrinse formulated from Aqueous Extract of *Terminalia chebula* on Salivary *Streptococcus mutans* Count and pH among 8- to 12-year-old School Children of Karnataka: A Randomized Clinical Trial

Madhuchanda Palit, Sundeep K Hegde, Sham S Bhat

ABSTRACT

**Aim:** The aim of the present study was to evaluate the anticariogenic efficacy of hot and cold aqueous extracts of *Terminalia chebula* against *Streptococcus mutans* as an oral rinse and also to discover the acceptability of the mouthwash in children.

**Settings and design:** Sixty children between 8 and 12 years with high caries risk were selected.

**Materials and methods:** 10% concentration of hot and cold aqueous extracts were prepared. Children were randomly divided into extract and control group. Baseline salivary samples were taken, and the samples were re-collected at 10, 60, and 90 minutes interval after rinsing. Microbial and pH analysis were done. An acceptability questionnaire was filled.

**Statistical analysis:** Tukey’s multiple comparison test.

**Results:** The results show statistically significant difference in *S. mutans* counts at 10, 60, and 90 minutes interval when compared with negative control. However, when the hot and cold extracts were compared, there was no significant difference. Acceptability questionnaire showed 65 to 75% overall acceptability for both types of extract.

**Conclusion:** Results of this study showed that both types of aqueous extract of *T. chebula* may be used as potential anticariogenic mouthwash with acceptable taste in children.

**Keywords:** Dental caries, Herbal mouth rinse, *Streptococcus mutans*, *Terminalia chebula*.

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INTRODUCTION

Dental caries is an irreversible microbial disease of calcified tissues of the teeth, characterized by demineralization of the inorganic portion and destruction of the organic substances of the tooth, which often leads to cavitation. Although there are effective methods known for prevention and management of dental caries, it is a major health problem affecting mankind. It is a complex and dynamic process where a multitude of factors influence and initiate the progression of disease. However, it is uniformly agreed that caries cannot occur without the presence of microorganisms. Evidences implicating its role in etiology of caries is elucidated as germ free animals do not develop caries, antibiotics administered to animals results in reducing the incidence of caries, unerupted teeth do not develop caries, and microorganisms have been demonstrated in enamel and dentinal caries.

*Streptococcus mutans* is considered the primary pathogen in dental caries development. This cariogenic organism is acquired early in life and the elimination or reduction of such pathogenic bacteria is beneficial in controlling dental caries. Because of this bacterial origin of dental caries, chemotherapeutic agents constitute a reasonable approach toward medical model of management of dental caries. These agents aid in the removal of microorganisms along with the mechanical plaque control, especially in children who are unable, unwilling, or untrained to practice routine effective mechanotherapy.

Ideal chemotherapeutic plaque control agent should have specificity, substantivity, stability, absence of adverse reactions, and easy to use. No agents have yet been developed that has all these characteristics. In contrary to this, many of the commonly used agents have the adverse effect of alteration of taste, burning sensation, staining, etc., which mandates the search for some newer agents. Plants or plant products are being used in folk dental practices or prescribed in Unani, homeopathic, or ayurvedic remedies since ages. The beneficial effects of plant products are also scientifically proved. *Terminalia chebula* is also a similar plant-derived medicine with numerous therapeutic effects. It is evident that the ripe fruit of *T. chebula*...
is valuable in the prevention and treatment of several
diseases of the mouth, such as dental caries, spongy and
bleeding gums, gingivitis, and stomatitis. Studies have
shown that tannic acid present in T. chebula is bacteriostatic
or bactericidal to some Gram-positive and Gram-negative
pathogens. Hence, this study aimed to evaluate the anti-
bacterial effect of two different types of aqueous extract of
T. chebula against S. mutans and its acceptability as mouth
rinse in children.

MATERIALS AND METHODS

This study was a single-blinded (microbiologist) random-
ized control trial conducted in the Department of Pedodon-
tics and Preventive Dentistry in collaboration with the
Research Center and Department of Microbiology after
obtaining Ethical Committee permission. Double blinding
could not be done, as it was necessary to inform parents
as well as children, in detail regarding the mouthwash
and its composition.

Preparation of the Extract

The dried ripe fruits of T. chebula were obtained and
ground to fine powder and 10% concentration of hot
and cold aqueous extracts were prepared. Cold aqueous
extract was prepared by suspending the ripe fruit (400 gm)
in 10 times its quantity of sterile distilled water in a round
bottomed flask and kept at 4°C for 72 hours. The aqueous
extract was then decanted, clarified by filtration through
a muslin cloth, and evaporated in a flat-bottomed por-
celain dish at 40°C. The dried extract was stored at 4°C
prior to use. The extract was suspended in polyethylene
glycol (PEG) 400 (20% v/v) and sterile distilled water to
give a final concentration of 30% w/v. The concentrated
extract was diluted with sterile distilled water to give
concentrations of 10% w/v.

The hot aqueous extract was freshly prepared by
adding 50 mg of T. chebula powder to 50 mL of sterile
distilled water and boiling to give a concentration of
10% w/v.

Both hot as well as cold aqueous extract were used as
mouth rinse at normal temperature. They differed only
in their process of making. Distilled water was used as
negative control for both the groups.

Study Population and Design

All the children coming to the Department of Pedodon-
tics for regular checkup were approached. The children
between 8 and 12 years were randomly assigned and
checked for the inclusion and exclusion criteria. Sixty
children were randomly divided into study and control
groups. Twenty children were selected as control and 20
each in both the study groups (Flow Chart 1). Information
regarding the nature of the study was explained to the
parents and children and an informed consent was
obtained. Only normal healthy children with full comple-
tment of teeth and 1 to 2 established caries lesion were
selected. High caries risk assessment was done using
AAPD Caries-risk Assessment Tool (CAT). Children with
any systemic diseases, on medication, or any intraoral
appliances/prostheses were considered to be the exclu-
sion criteria for the study. The children were randomly
divided into study and control group. They were not
allowed to eat 2 hours prior to the baseline sample collec-
tion. Five milliliters of hot and cold extract were given to the
assigned groups respectively 10 minutes after the baseline
sample collection and the children were asked to rinse for
1 minute. Salivary sample were collected again at 10, 60,
and 90 minutes after rinsing. Unstimulated saliva was
allowed to collect at the floor of the mouth and the saliva
was collected in two milliliter collection vial.

pH Analysis

Salivary pH analysis was done by a pH paper in the
range of 6 to 8.

Microbial Analysis

For assessing the microbial content, dilution and spread
plate was used. Salivary microbial analysis was done by
diluting each salivary sample 1:10 times. Each sample
was then streaked into Mitis salivarius agar media to
determine the total Streptococcus count, including the
S. mutans count. Plates were then incubated for 48 hours
at 37°C. The numbers of bacterial colonies were counted
in a colony counter to avoid errors.

Acceptability

An acceptability questionnaire was prepared and given
to all the children consisting of questions regarding
taste, color, odor, and overall acceptance of the extracts.
Effectiveness of Mouthrinse formulated from Aqueous Extract of Terminalia chebula on Salivary Streptococcus mutans


The questionnaire consisted of closed-ended questions regarding whether the extracts were pleasant, acceptable, or unacceptable with respect to color, taste, and odor. Overall acceptability was based on the acceptance of the extracts by the children in terms of color, taste, and odor.

For intragroup comparison of the paired sample, t-test was applied, while for the intergroup comparison between the three groups at different time intervals, Tukey’s multiple comparison tests were applied. All the tests were carried out using Statistical Package for the Social Sciences (SPSS) package in the computer. According to the answers provided by the study subjects, the results of the acceptability questionnaire were expressed in terms of percentage.

RESULTS

pH Analysis

When the baseline pH of both the extract groups were compared to the pH at 10, 30, and 90 minutes, there was an increase in pH and it remained alkaline for a period of 90 minutes after rinsing. For cold extract group the mean baseline pH was 6.65 and the mean pH at 90 minutes after the procedure was 7.80 (p < 0.001) (Table 1). For the hot aqueous extract group the mean baseline pH was 6.50 and 7.35 at the interval of 90 minutes after rinsing (p < 0.001) (Table 2). A comparison of pH at 10, 60, and 90 minutes but there was no statistically significant difference between the pH of hot and cold aqueous extract groups (p > 0.05) (Table 3).

Microbial Analysis

Microbial analysis of salivary samples indicated that there was reduction in salivary S. mutans colony at 10, 60, and 90 minutes after rinsing in both the extract groups compared with the baseline S. mutans colony count (p < 0.001) (Tables 4 and 5). When the hot and cold aqueous extract

<p>| Table 1: Intragroup comparison of baseline pH of cold aqueous extract to 10, 30, and 60 minutes (by paired t-test) |</p>
<table>
<thead>
<tr>
<th>Mean</th>
<th>Std. deviation</th>
<th>t-value</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pair 1</td>
<td>pH before 6.65</td>
<td>0.474</td>
<td>-11.000</td>
</tr>
<tr>
<td>pH 10 min 8.300</td>
<td>0.5375</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pair 2</td>
<td>pH before 6.65</td>
<td>0.474</td>
<td>-8.135</td>
</tr>
<tr>
<td>pH 60 min 7.90</td>
<td>0.516</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pair 3</td>
<td>pH before 6.65</td>
<td>0.474</td>
<td>-7.667</td>
</tr>
<tr>
<td>pH 90 min 7.80</td>
<td>0.537</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<p>| Table 2: Intragroup comparison of baseline pH of hot aqueous extract to 10, 30, and 60 minutes (by paired t-test) |</p>
<table>
<thead>
<tr>
<th>Mean</th>
<th>Std. deviation</th>
<th>t-value</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pair 1</td>
<td>pH before 6.50</td>
<td>0.527</td>
<td>-6.228</td>
</tr>
<tr>
<td>pH 10 min 7.750</td>
<td>0.5401</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pair 2</td>
<td>pH before 6.50</td>
<td>0.527</td>
<td>-6.678</td>
</tr>
<tr>
<td>pH 60 min 7.55</td>
<td>0.158</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pair 3</td>
<td>pH before 6.50</td>
<td>0.527</td>
<td>-3.791</td>
</tr>
<tr>
<td>pH 90 min 7.35</td>
<td>0.337</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<p>| Table 3: Intergroup comparison of pH of cold, hot aqueous extracts, and negative control at baseline, 10, 30, and 60 minutes (by Tukey’s multiple comparison test) |</p>
<table>
<thead>
<tr>
<th>Dependent variable</th>
<th>(I) group</th>
<th>(J) group</th>
<th>Mean difference (I–J)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>pH before</td>
<td>Cold extract</td>
<td>Hot extract</td>
<td>0.150</td>
<td>0.830</td>
</tr>
<tr>
<td>Hot extract</td>
<td>Negative control</td>
<td>0.250</td>
<td>0.600</td>
<td></td>
</tr>
<tr>
<td>pH 10 min</td>
<td>Cold extract</td>
<td>Hot extract</td>
<td>0.5500</td>
<td>0.094</td>
</tr>
<tr>
<td>Hot extract</td>
<td>Negative control</td>
<td>1.9000*</td>
<td>0.000</td>
<td></td>
</tr>
<tr>
<td>pH 60 min</td>
<td>Cold extract</td>
<td>Hot extract</td>
<td>0.350</td>
<td>0.283</td>
</tr>
<tr>
<td>Hot extract</td>
<td>Negative control</td>
<td>1.3500*</td>
<td>0.000</td>
<td></td>
</tr>
<tr>
<td>pH 90 min</td>
<td>Cold extract</td>
<td>Hot extract</td>
<td>0.450</td>
<td>0.174</td>
</tr>
<tr>
<td>Hot extract</td>
<td>Negative control</td>
<td>1.4000*</td>
<td>0.000</td>
<td></td>
</tr>
</tbody>
</table>

*Depicts that the mean differences are statistically significant

<p>| Table 4: Intragroup comparison of baseline S. mutans colony count of cold aqueous extract to 10, 30, and 60 minutes (by paired t-test) |</p>
<table>
<thead>
<tr>
<th>Mean</th>
<th>Std. deviation</th>
<th>t-value</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pair 1</td>
<td>S. mutans before 717.90</td>
<td>23.101</td>
<td>74.910</td>
</tr>
<tr>
<td>S. mutans 10 minutes 167.40</td>
<td>7.933</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pair 2</td>
<td>S. mutans before 717.90</td>
<td>23.101</td>
<td>114.532</td>
</tr>
<tr>
<td>S. mutans 60 minutes 195.50</td>
<td>14.057</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pair 3</td>
<td>S. mutans before 717.90</td>
<td>23.101</td>
<td>59.696</td>
</tr>
<tr>
<td>S. mutans 90 minutes 269.30</td>
<td>42.859</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<p>| Table 5: Intragroup comparison of baseline S. mutans colony count of hot aqueous extract to 10, 30, and 60 minutes (by paired t-test) |</p>
<table>
<thead>
<tr>
<th>Mean</th>
<th>Std. deviation</th>
<th>t-value</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pair 1</td>
<td>S. mutans before 674.90</td>
<td>35.534</td>
<td>39.461</td>
</tr>
<tr>
<td>S. mutans 10 min 170.80</td>
<td>8.162</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pair 2</td>
<td>S. mutans before 674.90</td>
<td>35.534</td>
<td>44.593</td>
</tr>
<tr>
<td>S. mutans 60 min 192.60</td>
<td>22.619</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pair 3</td>
<td>S. mutans before 674.90</td>
<td>35.534</td>
<td>29.271</td>
</tr>
<tr>
<td>S. mutans 90 min 233.20</td>
<td>38.169</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
groups were compared with the negative control, there was a lower S. mutans colony count at 10, 60, and 90 minutes (p < 0.001). However, when both the extract groups were compared at 10, 60, and 90 minutes, there was no difference in S. mutans colony count (p > 0.05) (Table 6).

### Acceptability

The results of acceptability questionnaire showed that out of the three categories of pleasant, acceptable, and unacceptable, almost all the subjects found the extracts acceptable with respect to color, odor, taste, and overall acceptability. None of the children found taste and overall acceptability of both the extract group pleasant. Out of the 20 subjects in each cold and hot extract group, color was acceptable to 18 (90%), odor to 17 (85%), taste to 17 (85%), and overall acceptability to 13 (65%) subjects respectively. Overall acceptability was 65 and 75% respectively for cold and hot extracts (Table 7).

### DISCUSSION

For thousands of years, medicinal plants have played an important role throughout the world in treating and preventing various diseases. T. chebula, also known as Haritaki, chebulic Myrobalan, or inknut, is a similar plant-derived medicine, which has been used since antiquity to control numerous human diseases. It is always listed first in the Ayurvedic Materia Medica in India. It exerts a wide range of pharmacological effects, including antibacterial, antiviral, antifungal, antioxidant, antiarthritic, antiaging properties.5,6,12-15 It also exhibits good hepatoprotection and nephroprotection against acetaminophen toxicity.16

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**Table 6: Intergroup comparison of S. mutans colony count of cold, hot aqueous extracts, and negative control at baseline, 10, 30, and 60 minutes (by Tukey’s multiple comparison test)**

<table>
<thead>
<tr>
<th>Dependent variable</th>
<th>(I) group</th>
<th>(J) group</th>
<th>Mean difference</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>S. mutans before</td>
<td>Cold extract</td>
<td>Hot extract</td>
<td>43.000</td>
<td>0.026</td>
</tr>
<tr>
<td>S. mutans 10 min</td>
<td>Hot extract</td>
<td>Negative control</td>
<td>13.400</td>
<td>0.666</td>
</tr>
<tr>
<td>S. mutans 60 min</td>
<td>Hot extract</td>
<td>Negative control</td>
<td>-29.600</td>
<td>0.154</td>
</tr>
<tr>
<td>S. mutans 90 min</td>
<td>Cold extract</td>
<td>Hot extract</td>
<td>-3.400</td>
<td>0.941</td>
</tr>
<tr>
<td>S. mutans 60 min</td>
<td>Hot extract</td>
<td>Negative control</td>
<td>-544.900</td>
<td>0.000</td>
</tr>
<tr>
<td>S. mutans 90 min</td>
<td>Hot extract</td>
<td>Negative control</td>
<td>-544.400</td>
<td>0.000</td>
</tr>
<tr>
<td>S. mutans 90 min</td>
<td>Cold extract</td>
<td>Hot extract</td>
<td>2.900</td>
<td>0.961</td>
</tr>
<tr>
<td>S. mutans 60 min</td>
<td>Hot extract</td>
<td>Negative control</td>
<td>-541.500</td>
<td>0.000</td>
</tr>
<tr>
<td>S. mutans 90 min</td>
<td>Cold extract</td>
<td>Hot extract</td>
<td>36.100</td>
<td>0.136</td>
</tr>
<tr>
<td>S. mutans 90 min</td>
<td>Hot extract</td>
<td>Negative control</td>
<td>-454.700</td>
<td>0.000</td>
</tr>
<tr>
<td>S. mutans 90 min</td>
<td>Hot extract</td>
<td>Negative control</td>
<td>-490.800</td>
<td>0.000</td>
</tr>
</tbody>
</table>

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**Table 7: Results of acceptability questionnaire**

<table>
<thead>
<tr>
<th>Acceptability</th>
<th>Cold aqueous extract (n = 20)</th>
<th>Hot aqueous extract (n = 20)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Color</td>
<td>18 (90%)</td>
<td>19 (95%)</td>
</tr>
<tr>
<td>Odor</td>
<td>17 (85%)</td>
<td>19 (95%)</td>
</tr>
<tr>
<td>Taste</td>
<td>11 (55%)</td>
<td>13 (65%)</td>
</tr>
<tr>
<td>Overall</td>
<td>13 (65%)</td>
<td>15 (75%)</td>
</tr>
</tbody>
</table>

Literature review reveals abundant evidence for the use of plants and plant products in preventing dental caries.17,18 In our study, we have selected T. chebula – to evaluate the efficacy of its extract as an anticariogenic mouth rinse in children.

Tannins are a group of polymeric phenolic compounds and they are the major constituent of T. chebula responsible for its antimicrobial property against S. mutans. Tannins inhibit enzyme glucosyltransferase by virtue of some tannin–protein interaction, hence affecting dextran-induced aggregation of S. mutans on the tooth surface. It also inhibits the acid production and sucrose-mediated adherence of S. mutans.5 In the present study, we have used an aqueous solution of T. chebula as most of the plant products are soluble in it. Methanol or ethanol extract could have given a better result, but the adverse effect of alcoholic extracts in children has further warranted us to use aqueous extracts.19

We used both hot as well as cold aqueous extracts at 10% concentration according to the works of Jagtap and Karkera.5 They suggested that at concentrations of 10%, the extract had an immediate effect on the salivary bacteria, and this effect was retained for 3 hours. However, at a lower concentration it could not inhibit S. mutans growth. Hence the rinsing of the mouthwash with water should be avoided, as it will cause dilution of the mouthwash and might decrease its efficiency by reducing the concentration.

The hot and cold aqueous extract groups differed in their preparation method. Hot extract was prepared according to the works of Nayak et al,8 while the methodology of Carounanidy et al7 was followed to prepare cold aqueous extract. Both the extracts, however, were used as a mouth rinse at normal temperature.

In hot aqueous extract group, the extract were freshly prepared, hence solvent was not needed; whereas in the cold aqueous extract group, PEG was used as solvent. Results of our study showed that both types of extracts at similar concentrations were equally effective in reducing S. mutans count. Hence, in the absence of pharmaceutical grade PEG, hot aqueous extract can also be used with equal effectiveness, however, the preparation of fresh extract every time will be cumbersome for larger study population.
Unstimulated salivary pH was analyzed using color indicator pH strips. Unstimulated saliva was used, as resting salivary pH plays a major role in caries initiation. Our study showed an increase in pH up to 90 minutes after rinsing. Nayak et al. and Carounanidy et al. noted an increase in pH up to 1 hour and 45 minutes. Jagtap and Karkera noted an increase at 2, 3, and 5 hours after rinsing. Hence, both the extracts have a high substantivity, which is essential for an effective mouthwash.

The present study showed a statistically significant reduction in S. mutans colony count at 10 minutes after rinsing with both types of aqueous extracts and this decrease in the colony remained below the baseline even after 90 minutes. Similar sustained decrease in salivary bacteria for a period of 3 hours was noted by Jagtap and Karkera. According to Nayak et al., there was a observed reduction in the colony count up to 1 hour after rinsing. This reduction is mainly because tannic acid binds strongly to the carboxyl groups of the salivary glycoproteins present on the pellicle and hence prolonging its antimicrobial activity against S. mutans.

For counting the S. mutans colony, we have used 1:10 dilution, whereas Nayak et al. have used 1:100 dilutions. Decreasing the dilution (1:10) as done in our study will give a more crowded colony as compared with higher dilution (1:100), which might be the reason for the difference in mean between the two studies. Since the crowded colony would be difficult to count, we have used a microbial colony counter, which has as well reduced the error.

Salivary S. mutans colony count and pH were evaluated at different time intervals to evaluate its effect for a prolonged time. The result of our study showed that there was definite decrease in all the parameters even after 90 minutes as compared with baseline, but there was a gradual increase at 10, 60, and 90 minutes (Graphs 1 and 2). Hence, further studies for a longer duration might help us to determine the dose interval of the mouth rinse, i.e., how frequently it should be prescribed to obtain a sustained reduced oral S. mutans count.

To measure the acceptability of both the hot and cold aqueous extracts, we have given closed-ended questionnaire to the children. The extracts were categorized as “pleasant,” “acceptable,” or “unacceptable” in terms of “color,” “odor,” “taste,” and “overall acceptability.” In our study, result of the acceptability questionnaire indicated that the color, odor, and taste was acceptable at 90, 85, and 55% for cold extract, whereas for hot extract it was 95, 95, and 65% respectively. Majority of the children found taste, color, and odor as “acceptable.” The overall acceptability was also “acceptable” to most (65% for cold and 75% for hot extract) of the children. As only 5 children out of 40 for both the extract groups found overall acceptability as “pleasant” and none of them found it “unacceptable,” the overall acceptability for the categories “pleasant” and “unacceptable” was not considered. These results were similar to the results by Nayak et al. Hence, the overall acceptability results indicated that the extracts may be acceptable to most children.

Ours is the first study where T. chebula mouth rinse has been used in children. We have studied the effects of both types of T. chebula aqueous extract; a comparison study of the rinse over conventionally available mouth rinses was not done. Hence, further studies are required to compare the aqueous extracts to the conventionally available mouth rinses in children.

The present study is limited to a short duration of 90 minutes. Further research should also focus on long-term use of the mouth rinses on larger sample size as well as dose determination.

T. chebula may therefore be used as an anticariogenic mouthwash in children due to its effectiveness in reducing oral load of S. mutans. It is economical, easy to prepare,
has good substantivity, and also provides multiple beneficial health effects. It may also be well accepted by the children, however long-term study with larger sample size and dose determination is further required.

**Importance to Pediatric Dentistry**

- *T. chebula* may be used as an effective oral rinse in children with high caries risk to reduce the oral load of *S. mutans*.

**REFERENCES**